

1 **Association between polymorphisms in adiponectin, leptin and leptin**
2 **receptor genes with COVID-19 disease severity among Jordanian**
3 **population**

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7 **Running title:** Adipokines genetic variations and COVID-19 disease

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25 **Abstract**

26 **Introduction:** Variations in genetic makeup can affect COVID-19 disease
27 susceptibility and severity. Adipokines, including leptin and adiponectin, are protein
28 hormones synthesized in the adipose tissues. Adipokines are implicated in the
29 regulation of body metabolism and immunity. Here, the study investigated the
30 relationships between single nucleotide polymorphisms in adiponectin (*ADIPOQ*:
31 rs1501299 and rs822396), leptin (*LEP*: rs7799039) and leptin receptor (*LEPR*:
32 rs1137101) and COVID-19 susceptibility/severity in Jordan.

33 **Materials and Methods:** A cross-sectional, case-control design was adopted. The
34 study included 427 subjects who never experienced COVID-19 disease (controls), and
35 434 subjects with COVID-19 disease history (cases). Cases included 268 subjects
36 who experienced mild/moderate symptoms and 166 subjects who experienced severe
37 symptoms. Genotyping of genetic variants was performed using “polymerase chain
38 reaction-restriction fragment length polymorphism technique (PCR-RFLP)”
39 technique. Cross-tabulation and multivariate regression analysis were used in data
40 analysis.

41 **Results:** No statistical differences were found between distribution of genotypes of
42 the studied genetic variations and susceptibility to COVID-19 ($P>0.05$). Regarding
43 disease severity, the *ADIPOQ* rs1501299 polymorphism exhibited an association with
44 COVID-19 severe form of the disease ($P<0.05$). The frequency of GT genotype was
45 lower ($P<0.05$) among patients with a severe form of the disease than among the
46 patients with a non-severe form of the disease. Finally, the AGGA haplotype of
47 rs822396, rs1501299, rs1137101, and rs7799039 polymorphisms was higher among
48 patients with severe form of the disease than among patients with non-severe form of
49 the disease ($P<0.01$).

50 **Conclusion:** the *ADIPOQ* rs1501299 polymorphism and the AGGA haplotype of
51 rs822396, rs1501299, rs1137101, and rs7799039 may contribute to COVID-9 disease
52 severity among Jordanian people. The current results can be integrated in the
53 management of COVID-9 disease severity.

54 **Keywords:** adipokines, genetic variations, SARS-CoV-2, virus

55

56 **1. Introduction**

57 COVID-19 is a serious contagious infection caused by “severe acute respiratory
58 syndrome coronavirus (SARS-CoV-2)”. Symptoms range from asymptomatic
59 infection to a life-threatening complicated pneumonia [1]. The virus invades various
60 tissues in the body, primarily the respiratory system, and then spreads to other body
61 systems such as the neuronal system, gastrointestinal tract, and circulatory system [2].
62 Symptoms include sore throat, fever, hypoxia, dyspnea, diarrhea, and others. The
63 variability of disease symptoms is attributed to several factors, including age, health
64 status, and genetic factors [1].

65 Adipokines play a role in regulating body’s metabolism and immunity [3]. The most
66 important adipokines are adiponectin and leptin [4]. Adiponectin is the product of
67 *ADIPOQ* gene in the adipose tissue that circulates in the bloodstream to the target
68 cells. Adiponectin has anti-inflammatory and antioxidant properties and enhances
69 insulin sensitivity in the human body [5]. Like adiponectin, leptin is a protein
70 produced by adipose tissues and circulates in the bloodstream. It is encoded by *LEP*
71 gene and involved in regulating appetite, energy expenditure, and immune response
72 [6]. Some investigations have examined the relationship between COVID-19 and
73 adiponectin and leptin blood concentrations, with conflicting results [7]. Most studies
74 agree on an inverse relationship between circulating adiponectin concentration and the

75 degree of COVID-19 severity [8, 9]. Regarding leptin, elevated adiponectin levels are
76 associated with an excessive inflammatory response and the resulting severe form of
77 COVID-19 [8, 10, 11]. In addition, adipose tissue expresses ACE2, the major receptor
78 for the “SARS-CoV-2”.

79 Adiponectin and leptin levels are impacted by “single nucleotide polymorphisms”
80 (SNPs). These include rs822396 (-3971A>G) and rs1501299 (276G>T) in *ADIPOQ*
81 gene, rs7799039 (2548A>G) in *LEP* gene, and rs1137101 (A>G: Q223R) in leptin
82 receptor (*LEPR*) gene. The rs822396 and rs1501299 SNPs influence the expression
83 level of adiponectin and have been reported to affect the risk to several long-lasting
84 illnesses such as metabolic syndrome, diabetes, and asthma [12]. Similarly, rs7799039
85 in the *LEP* gene influences leptin levels and is linked to diabetes, obesity, and cancer.
86 The rs7799039 has also been shown to affect COVID-19 severity in the Egyptian
87 population [13]. Finally, rs1137101 in the *LEPR* gene is an important polymorphism
88 that impacts leptin function and has been shown to be associated with several chronic
89 diseases . Therefore, the aim of this investigation was to study the contribution of
90 rs822396, rs1501299, rs7799039, and rs1137101 to susceptibility/severity of COVID-
91 19 disease in Jordan.

92 **2. Materials and Methods**

93 **2.1. Study Subjects**

94 The “case-control design” was adopted in the current investigation. The case group
95 included 434 participants who previously infected with COVID-19. The control group
96 included 427 participants without a history of COVID-19. Both groups were recruited
97 from northern Jordan and were age- and sex- matched. COVID-19 history was
98 confirmed through participants’ medical records. Cases were stratified into: non-

99 severe and severe subgroups, using the severity scale described previously [14]. Non-
100 Jordanians and children (<18 years old) were not allowed to participate.

101 **2.2. Ethical issues**

102 The “Institutional Review Board (IRB) of Jordan University of Science and
103 Technology” approved the study protocol (approval ID: 20/160/2023). Participants
104 were given written informed consent in accordance with IRB and international
105 research ethical regulations.

106 **2.3. Blood Sampling**

107 Blood samples were drawn from participants in EDTA blood collection tubes. Blood
108 was stored in the freezer for DNA extraction and genotyping.

109 **2.4. Anthropometric Measurements**

110 Participant's height (H) was measured using a stadiometer, while their weight (W)
111 was obtained using a medical scale. BMI was deducted from W(kg) and H(m²) as
112 previously described. Body fat percentage was measured using a smart medical scale
113 obtained from ICOMON (China).

114 **2.5. Molecular Analysis**

115 DNA was extracted using a “G-spin™ genomic DNA extraction kit” (iNtRON
116 Biotech) as described in the kit datasheet. DNA quantity and quality were measured
117 using a nanodrop spectrophotometer (Bio-Rad, USA).

118 Genotypes of rs7799039, rs1137101, and rs822396 variants were detected via
119 “polymerase chain reaction-restriction fragment length polymorphism technique”.
120 Amplification conditions and restriction conditions are shown in Table 1. After PCR-
121 RFLP procedure, the digestion products were electrophoresed on 3% agarose,
122 visualized using EtBr under UV light.

123

124 **Table 1.** PCR and restriction conditions of examined polymorphisms of included
 125 genes.

| Item | rs7799039 | rs1137101 | rs822396 |
|------------------------|---|--|---|
| F primers | 5'TTTCTGTAATTTTCC CGTGAG3' | 5'GGCCTGAAGTGTT AGAAGAT3' | 5'GGTCTTGGAAAC TTCTGAGGCT3' |
| R primers | 5'AAAGCAAAGACAGG CATAAA3' | 5'CTGCTCTCTGAGG TGGGAAC3' | 5'AAACCTTGGAG AGAGGGCAA3' |
| PCR product | 242 bp | 642 bp | 245 bp |
| PCR conditions | 94°/5min then 34 cycles: 94°C/30s, 52°C/45s, and 72°C/45s, then a final extension of 72°C/5min | 94°/5min then 32 cycles: 94°C/30s, 58°C/30s, and 72°C/30s, then a final extension of 72°C/5min | 94°/3min then 35 cycles: 94°C/30s, 55°C/25s, and 72°C/30s, then a final extension of 72°C/5min |
| Restriction conditions | 10 units of <i>HhaI</i> /10uL of PCR product incubated at 37°C overnight | 10 units of <i>MspI</i> /10uL of PCR product incubated at 37°C overnight | 10 units of <i>MseI</i> /10uL of PCR product incubated at 37°C overnight |
| Restriction products | AA: 242bp GG: 181/61bp | AA: 642bp GG: 469/173bp | GG: 245bp, AA: 181 + 64 |

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128 The genotypes of *ADIPOQ* rs1501299 were determined using the polymerase chain
 129 reaction-allele refractory mutation system (PCR-ARMS). Amplification of region
 130 flanking rs1501299 was performed using forward
 131 (5'GAGCTGTTCTACTGCTATTAGCTCTGC3') and reverse primers
 132 (5'GAATATGAATGTACTGGGAATAGGGATG3'). Internal forward primer for the
 133 G allele was: 5'CCTCCTACACTGATATAAACTATATGAGGG'3 and the internal
 134 reverse primer for the T allele was:
 135 5'TGTGTCTAGGCCTTAGTTAATAATGAACGA3'. PCR conditions were
 136 95°/5min then 32 cycles (95°C/0.5min, 58°C/0.5min, and 72°C/1min), followed by
 137 72°C/5min [15]. After PCR, fragments were electrophoresed on 3% agarose,
 138 visualized using EtBr under UV light. The PCR fragments were: 476bp for the
 139 "control band", 244bp for the "G allele", and 292bp for the "T allele".

140 **2.6. Statistics**

141 The “SNPStats” program was utilized to determine rs7799039, Q223R, rs822396, and
 142 rs1501299 distribution and relationship with susceptibility/severity of COVID-19.
 143 The alignment of the examined polymorphisms with Hardy–Weinberg equilibrium
 144 was also assessed using “SNPStats”. Study measures are presented as mean (\pm SD) or
 145 percentages, and they were compared between groups using SPSS (version 22). The
 146 cutoff p-value for significant was ≤ 0.05 . Sample size was computed using “GPower-
 147 3.1 software” (intermediate effect/ $\alpha=0.05/1-\beta=0.9$). The minimum required
 148 sample=470 participants.

149 **3. Results**

150 The study included 434 participants with a history of COVID-19 disease (cases) and
 151 427 subjects without a history of COVID-19 disease (controls). No statistical
 152 difference ($P > 0.05$) was observed between cases and controls regarding age, and
 153 gender ratios. However, differences were found between cases and controls
 154 concerning BMI, body fat percentage, and the presence of chronic diseases ($P < 0.05$,
 155 Table 2). The incidence of chronic diseases and body fat percentage were
 156 significantly higher among the case group, while BMI was slightly higher among the
 157 control group.

158 **Table 2:** Demographic Characteristics of the overall subjects:

| Variables | Case (n=434) | Control (n=427) | P-value |
|------------------------------------|---------------------|------------------------|----------------|
| Gender ratio (male: female) | 41.9:58.1 | 46.1:53.9 | 0.210 |
| Age (years) | 46.0 \pm 13.4 | 47.2 \pm 12.3 | 0.180 |
| BMI (kg/m²) | 30.4 \pm 5.8 | 31.5 \pm 8.3 | <0.05 |
| % body fat | 36.6 \pm 9.24 | 32.4 \pm 8.33 | <0.01 |
| Having a chronic disease | 171(39.4%) | 139(32.6%) | <0.01 |

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161 The distribution of genotypes of the included variants among participants is shown in

162 Table 3. The AA genotype (71.7%) of rs822396, the GG (42.6%) genotype of the

163 rs1501299, AG genotype (49.1%) of rs7799039, and AA genotype of rs1137101 were
 164 common in the study sample. None of the examined polymorphisms were associated
 165 ($P>0.05$) with COVID-19 susceptibility.

166

167 **Table 3:** Genotype distribution of rs822396 based on susceptibility to COVID-19

| Polymorphism | Genotype | Case | Control | OR (95% CI) | P-value |
|--------------|----------|------------|------------|------------------|---------|
| rs822396 | AA | 311 (71.7) | 334 (78.2) | 1.00 | 0.068 |
| | AG | 113 (26) | 83 (19.4) | 1.46 (1.06-2.02) | |
| | GG | 10 (2.3) | 10 (2.3) | 1.07 (0.44-2.62) | |
| rs1501299 | GG | 214 (49.3) | 182 (42.6) | 1.00 | 0.096 |
| | GT | 158 (36.4) | 185 (43.3) | 0.73 (0.54-0.97) | |
| | TT | 62 (14.3) | 60 (14.1) | 0.88 (0.59-1.32) | |
| rs1137101 | AA | 209 (48.2) | 202 (47.3) | 1.00 | 0.97 |
| | AG | 182 (41.9) | 182 (42.6) | 1.03 (0.78-1.37) | |
| | GG | 43 (9.9) | 43 (10.1) | 1.03 (0.65-1.65) | |
| rs7799039 | GG | 122 (28.1) | 102 (23.9) | 1.00 | 0.323 |
| | AG | 213 (49.1) | 228 (53.4) | 1.28 (0.93-1.77) | |
| | AA | 99 (22.8) | 97 (22.7) | 1.17 (0.80-1.72) | |

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170 Next, we examined the association between rs822396, rs1501299, rs7799039,
 171 rs1137101 haplotypes and susceptibility to COVID-19. Similar to genotypes, none of
 172 the haplotypes were associated with COVID-19 susceptibility.

173 The case group was stratified into two categories according to COVID-19 severity. As
 174 shown in Table 4, the *ADIPOQ* rs1501299 polymorphism was associated with severe
 175 form of COVID-19. The frequency of GT genotype was lower among the severe
 176 subjects compared to non-severe ones. However, rs822396, rs7799039, and
 177 rs1137101 did not impact COVID-19 severity ($P>0.05$).

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181 **Table 4:** Genotype distribution of adiponectin polymorphisms within the case
 182 subgroups based on COVID-19 severity.

| Polymorphism | Genotypes | Severe | Non-severe | OR (95% CI) | P-value |
|--------------|-----------|------------|------------|------------------|---------|
| rs822396 | AA | 117 (70.5) | 194 (72.4) | 1.00 | 0.730 |
| | AG | 46 (27.7) | 67 (25.0) | 1.14 (0.73-1.77) | |
| | GG | 3 (1.8) | 7 (2.60) | 0.71 (0.18-2.80) | |
| rs1501299 | GG | 91 (54.8) | 123 (45.9) | 1.00 | 0.038 |
| | GT | 48 (28.9) | 110 (41.0) | 0.59 (0.38-0.91) | |
| | TT | 27 (16.3) | 35 (13.1) | 1.04 (0.59-1.84) | |
| rs7799039 | AA | 46 (27.7) | 53 (19.8) | 1.00 | 0.160 |
| | GA | 77 (46.4) | 136 (50.8) | 1.04 (0.65-1.66) | |
| | GG | 43 (25.9) | 79 (29.5) | 1.59 (0.93-2.74) | |
| rs1137101 | AA | 71 (42.8) | 138 (51.5) | 1.00 | 0.210 |
| | AG | 77 (46.4) | 105 (39.2) | 1.43 (0.95-2.15) | |
| | GG | 18 (10.8) | 25 (9.3) | 1.40 (0.72-2.74) | |

183

184 The association between rs822396, rs1501299, rs7799039, rs1137101 haplotypes and
 185 COVID-19 severity are presented in Table 5. The frequency of AGGA haplotype was
 186 higher among the “severe group” (0.126%) than among the “non-severe group”
 187 (0.046%, OR [95%CI]: 2.95 [1.41 - 6.16], P<0.01). On the other hand, none of the other
 188 haplotypes were significantly associated with COVID-19 severity.

189

190 **Table 5.** Haplotype analysis of rs822396 (A/G), rs1501299 (G/T), rs1137101 (G/A), and
 191 rs7799039 (A/G), polymorphisms and COVID-19 disease severity.

| | Non-severe | Severe | OR (95% CI) | P-value |
|------|------------|--------|---------------------------|-----------------|
| AGAG | 0.22 | 0.2075 | 1 | --- |
| AGAA | 0.1723 | 0.1579 | 0.95 (0.52 - 1.75) | 0.86 |
| ATAG | 0.1124 | 0.0971 | 0.97 (0.49 - 1.89) | 0.92 |
| ATAA | 0.112 | 0.0882 | 0.87 (0.46 - 1.65) | 0.67 |
| AGGG | 0.0974 | 0.0758 | 0.75 (0.33 - 1.70) | 0.49 |
| AGGA | 0.0462 | 0.1264 | 2.95 (1.41 - 6.16) | <0.01 |
| ATGG | 0.051 | 0.0595 | 1.18 (0.47 - 2.96) | 0.72 |
| GGAA | 0.0574 | 0.0536 | 1.08 (0.44 - 2.66) | 0.87 |
| ATGA | 0.0377 | 0.0311 | 0.86 (0.22 - 3.36) | 0.83 |
| GGAG | 0.0368 | 0.024 | 0.63 (0.15 - 2.71) | 0.54 |
| GGGA | 0.0152 | 0.0206 | 1.86 (0.48 - 7.12) | 0.37 |
| GTAA | 0 | 0.0313 | 1.21 (0.27 - 5.40) | 0.81 |
| GGGG | 0.019 | 0.0271 | 3.82 (0.22 - 65.75) | 0.36 |
| GTGG | 0.012 | 0 | 0 | - |
| GTGA | 0.0107 | 0 | 0 | - |

192 **4. Discussion**

193 COVID-19 can be expressed in different forms with a range of clinical signs. Previous
194 literature indicates that certain variants in genes can impact individual susceptibility
195 to COVID-19 and its severity. Here, the relationship between *ADIPOQ* (rs1501299
196 and rs822396), *LEP* (rs7799039) and *LEPR* (rs1137101) variants with COVID-19
197 susceptibility/severity was examined. No associations were observed between
198 genotypes of the examined variants and susceptibility to COVID-19. When disease
199 severity was considered, the *ADIPOQ* rs1501299 was found to impact COVID-19
200 severity. Additionally, AGGA haplotype of rs822396, rs1501299, rs1137101, and
201 rs7799039 was found to be associated with disease severity.

202 Adipokines are a group of proteins produced by adipose tissues and circulate in the
203 bloodstream. Adipokines such as adiponectin and leptin, are implicated in the
204 regulation of body metabolism and immunity [5]. Levels of these cytokines have been
205 shown to be associated with the etiology of many disease conditions, such as obesity,
206 diabetes, heart disease and cancer [12]. These conditions have been shown to
207 modulate outcomes of COVID-19 infection. In addition, circulating adiponectin and
208 leptin levels have been shown to impact severity and outcomes of COVID-19 [16-18].
209 The present results showed no impact for *ADIPOQ*, *LEP*, and *LEPR* variants on
210 susceptibility to COVID-19. Furthermore, none of haplotypes of examined
211 polymorphisms were associated with susceptibility to COVID-19. The limited
212 literature addressing genetic variations in adipokine genes and COVID-19
213 susceptibility make comparisons with literature inapplicable. Adiponectin receptors
214 (such as *adipoR1* and *adipoR2*) are expressed in various parts of the respiratory
215 system including epithelial cells, smooth muscle cells and lung macrophages.
216 Similarly, leptin receptor is expressed in lung epithelial cells and airway muscle cells.

217 A previous study showed that leptin did not involve in “SARS-CoV-2” infection
218 ability in laboratory models of lung infection [19]. Additionally, adiponectin level was
219 reported to be low in COVID-19 patients [20]. The absence of a relationship between
220 the examine SNPs and COVID-19 susceptibility suggests that these adipokines may
221 not be involved in binding to or entrance of “SARS-CoV-2” into body cells.

222 The present results reported an impact of *ADIPOQ* rs1501299 and COVID-19
223 severity. Additionally, AGGA haplotype of rs822396, rs1501299, rs1137101, and
224 rs7799039 was found to be associated with disease severity. Limited literature has
225 examined the relationship of adipokine gene polymorphisms with COVID-19 disease
226 severity. A previous study in Egypt showed an impact for *LEP* rs7799039 on COVID-
227 19 disease severity [13]. A study that was conducted in Iraq reported no relationship
228 between *ADIPOQ* rs266729 variant and COVID-19 disease severity [21]. On the
229 other hand, the relationship between adiponectin and leptin blood levels and the
230 severity of COVID-19 has been well investigated, and studies have shown conflicting
231 results. Regarding adiponectin, literature has indicated an association between high
232 [22], low [23, 24] or unchanged [25] adiponectin levels in severe form of COVID-19.
233 Regarding leptin, literature has indicated an association between high [8, 10, 11, 24],
234 low [23] or unchanged [25] leptin levels in severe form of COVID-19. A study
235 showed that gender and underlying diseases influence the relationship between
236 adiponectin protein level and severe form of COVID-19 [26]. In addition, it has been
237 shown that adiponectin/leptin ratio influences COVID-19 disease severity, rather than
238 individual level of adiponectin or leptin [27]. Additionally, blood adiponectin level at
239 hospital admission was inversely related to death and lung dysfunction in COVID-19
240 [28]. The consensus in literature is that severe COVID-19 is related to lower
241 circulating adiponectin level and higher circulating leptin level.

242 Among the study limitations is that it was conducted in Jordan, therefore, it is
243 recommended to expand its scope to include other populations in other countries. In
244 addition, circulating adiponectin and leptin levels were not investigated, and deceased
245 patients were not included due to adopted retrospective approach.

246 **5. Conclusion**

247 The *ADIPOQ* rs1501299 polymorphism may contribute to COVID-9 disease severity
248 in the Jordanian population. Additionally, the AGGA haplotype of SNPs rs822396,
249 rs1501299, rs1137101, and rs7799039 may be associated with disease severity. These
250 findings support the relationship between circulating adiponectin level and leptin level
251 with COVID-19 disease severity.

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254 Technology” for its support.

255 **Ethics**

256 Approval of the study protocol was granted from “Institutional Review Board of
257 Jordan University of Science and Technology”.

258 **Authors’ Contribution**

259 **ZO:** “Methodology, investigation, data curation, writing-original draft”.

260 **OK:** “Methodology, supervision, funding acquisition, writing-review and editing”.

261 **Conflict of Interest**

262 Authors have no “conflict of interest” to declare

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266 **Availability of data**

267 “Data are available on reasonable request from the corresponding author”.

268

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