

# *Neospora caninum* Infection in Rodents: A Molecular Study in Dairy Cattle Farms in Arak, Iran

Fatemeh Arabkhazaeli<sup>1\*</sup>, Mohammad Khani<sup>2</sup>, Seyed Davood Hosseini<sup>2</sup>, Mobina Farrokhnia<sup>3</sup>

<sup>1</sup>Department of Parasitology, Faculty of Veterinary Medicine, University of Tehran, Tehran, Iran

<sup>2</sup>Razi Vaccine and Serum Research Institute, Arak, Iran

<sup>3</sup>Faculty of Veterinary Medicine, University of Tehran, Tehran, Iran

## \* Correspondence:

Corresponding Author:

Fatemeh Arabkhazaeli

Qarib St., Azadi St., Department of Parasitology, Faculty of Veterinary Medicine, University of Tehran, Tehran, Iran

Tel.: +982161117049

Fax: +982166933222

Email: farab@ut.ac.ir

## Abstract

*Neospora caninum* is an apicomplexan protozoa which is an important cause of abortion and economic loss in dairy and beef industries. This parasite has an indirect prey-predator lifecycle, which provides an opportunity for domestic and wild species to play a role in the lifecycle of *N. caninum*. Ongoing research is being conducted to ascertain the involvement of other vertebrates in the epidemiology and transmission of this parasite. Rodents are abundant in many habitats, including livestock farms, and their role in the maintenance and spread of *N. caninum* remains unresolved. In this study the plausible role of feral rodents in the transmission of *N. caninum*, was investigated in wild rodents captured from several dairy farms with a history of abortion and neosporosis in Arak city, Iran. During the study, rodent samples were collected from 14 farms with high abortion rate. All the trapped rodents were identified as *Mus musculus*. The rodents were necropsied and the brain samples were tested by Nested-PCR. No evidence for *N. caninum* infection was detected in any of the rodents' samples.

**Keywords:** *Neospora caninum*, Dairy cattle, Abortion, Molecular biology, *Mus musculus*

## 1. Introduction

*Neospora* sp. is a protozoan parasite causing abortion and reduced fertility in animals specially in cattle. The wide range of intermediate hosts makes the parasite widely distributed around the world (2, 3). Neosporosis causes sporadic abortions and abortion storms on farms causing severe reproductive and economic losses in the cattle. Besides the parasite is responsible for neuromuscular disease in dogs around the world (2,4,5). The vertical transmission of *Neospora*, in addition to the horizontal transmission through oocyst ingestion, plays an important role in the maintenance and spread of the infection within a cattle herd (5,6). The established role of dogs and certain wild canids as the definitive hosts in the lifecycle and prevalence of *N. caninum* is widely recognized. Different serological tests including enzyme-linked immunosorbent assays (ELISAs), indirect fluorescent antibody testing (IFAT), and agglutination tests in addition to the PCR-based (polymerase chain reaction) methods can be used to diagnose the infection. Small animals were incriminated to the sylvatic cycle of the infection. Rodents play an important role in the transmission of various microorganisms and they are criticized to have a role in the complex lifecycle of *N. caninum* in cattle farms. Studies on the role of rodents in the epidemiology of neosporosis has revealed the infection in different rodent species with varying relative frequency from Zero to 40% (3,7-12). The prevalence rate of cattle neosporosis is reported 23.6% and 20% in Iran and other countries, respectively (13). Globally, a prevalence rate of 5% among rodents has been reported, while this rate is 16% in Iran.

The data on the presence and prevalence of *Neospora* infection is sparse and ongoing. Various bird and rodent species were reported to harbor the parasite reservoir (3,7,9). These infested animals may play an important role in the epidemiology of the disease as their infected tissues may be the source of the infection for other hosts in the parasite's lifecycle. This study was performed to further investigate the plausible role of feral rodents on the distribution and infection of neosporosis. For this aim, wild rodents captured from several dairy farms with a history of neosporosis and abortion in Arak city, Iran were investigated molecularly for the presence of *N. caninum*

## 2. Methods

This study took place on the dairy farms in Arak (34°05'30.26"N 49°41'20.98"E), a county in Markazi province, Iran. Sampling from dairy cattle farms with a history of abortion due to neosporosis (14) was done from around the fodder barn, the manger, the watershed, milking parlor and outdoor area. Regarding 95% confidence level, 5% margin of errors and 4% population proportion, the least sample size was determined to be 60. Wooden traps and mouse glue trap were used for sampling. The trapped mice were euthanized with ether, identified morphologically and necropsied for obtaining fresh brain samples (12). The ethical approval for this study was obtained from the ethics committee of the Faculty of Veterinary Medicine, University of Tehran (28864/6/2). Upon necropsy any visible clinical lesions were recorded and brain was excised, homogenized aseptically with PBS (pH=7.4).

The obtained samples were centrifuged at 21500xg for 5 minutes and DNA extraction steps were followed on the sediment by DNA extraction Kit (Cinnaclone, Iran) (15). The samples were tested for the presence of *Neospora* using Nested-PCR. Primers for NC-5 gene were applied using NC-6, NC-21, NC-7 and NC-10 as nested-PCR (16-18). The PCR reaction contained 0.2  $\mu$ M of each primer, 200  $\mu$ M of each dNTP, and 1.5 mM of MgCl<sub>2</sub>, 2.5 U of Taq DNA polymerase, and 2  $\mu$ l of DNA template in a total volume of 25  $\mu$ l. For each set of PCR amplification, *N. caninum* isolates as internal positive control, and reaction without DNA template as the negative control were included. The thermal cycler PCR program was as following: pre-denaturing 5 min at 94 °C; 94 °C for 30 s in 40 cycles of, 63 °C for 30 s and 72°C for 1 min and a final 5 min extension at 72 °C. The amplified PCR products were

visualized on 1.5% Agarose gel pre-stained with Nancy-520 DNA Gel Stain (Sigma-Aldrich, Dorset, UK) under UV light and gel images were recorded by Gel document.

### 3. Results and Discussion

Totally, 68 rodents were captured from 14 farms with high abortion rate. All the captured rodents were identified as *Mus musculus*. None of the CNS samples was positive neither via PCR nor in the nested PCR tests.

Several investigations have explored the involvement of rodents in the epidemiology of *Neospora caninum*. In the West Indies (16) PCR revealed an infection rate of 8.6% while serology showed an infection rate of 5.1% in *Mus musculus*. 13.8% infection rate was reported by PCR in Italy in *Mus musculus* (19). In Argentina, there have been reports of infection in *Mus musculus* by IFAT (0.8%) (20). The Netherlands has reported 15.4% infection rate in house mice (15). In Mexico, PCR indicates a high infection rate of 77%, while immunohistochemistry shows a rate of 15% (12). The Czech Republic–German hybrid zone reports a 3.6% infection rate (21). Studies in Iran used agglutination, IFAT, PCR and nested-PCR tests and reported various rodent species to be infected with an infection rate of zero to 31.9% (10,13). In the present study no infection was detected in the samples.

There are several reports where the parasite was not identified in rodents' samples. Fernández-Escobar et al. (11) conducted a study and found no presence of *N. caninum* in house mice, although they did report a prevalence of 1.3% in other micromammals such as rats, shrews, and other species of mice. Similarly, Nazari et al. (23) used molecular methods to examine urban rodents and were unable to detect *N. caninum*, but 39% of the samples tested positive using IFAT (10). Macháčová et al. (24) reported a serologic prevalence of 0.4% in 621 captured wild mammals, but no positive results were found in the captured house mice. These disparities could potentially arise from the examined organs. While some studies reported liver as the best target organ for *Neospora* detection in rodents, others have implied that brain or heart samples suits better (11, 25).

Prevalence data analysis requires special attention due to the constraints encountered due to the applied detection techniques. According to Jenkins (16), when focusing on the ITS loci a higher number of positive outcomes could be obtained in samples obtained from dairy farm environments, than Nc5 PCR. Besides there are reports that failed to confirm the molecularly detected infection through immunohistochemistry (2, 11, 22).

Regarding the sampling habitat, rodents residing in dry-land habitats were found to have a higher likelihood of being infected with *N. caninum* compared to those trapped in different habitats such as forests, rain-fed lands. Rodents inhabiting cattle farms with *N. caninum* abortion were more frequently infected than in peri-urban areas (20, 25, 26).

Diverse detection techniques, different sampling locations in relation to proximity to cattle farms, various rodent species, and examination of various organs may all contribute to the result diversification. It is worth mentioning that PCR or serology are commonly used detection methods to identify *N. caninum* infections in various animal species. These methods detect parasite DNA or specific antibodies in the host, but they do not necessarily indicate a viable or successful infection (27). However, the involvement of other animal species including rodents and birds in the maintenance of the parasite is still under study. It has been proven that pigeons and gerbils are the most susceptible hosts (2).

Although the presence of antibodies or the parasite's DNA in animals other than bovids and canids may make these animals a plausible host, it has not been proven in experimental studies. Despite the susceptibility of *Mus musculus* to infection, the role played by in the urban cycle of *N. caninum* infection appears to be negligible. It may be noted that the present study surveyed brain tissue from the farm captured *Mus musculus*. In order to surpass the limitations on experiment parameters, it is recommended to incorporate multiple diagnostic and confirmatory

134 techniques on different organs, alongside a more extensive sampling approach that  
135 encompasses a wider range of rodent species.

### 136 **Acknowledgments**

137 The authors would like to thank our colleagues in the faculty of Veterinary Medicine,  
138 University of Tehran and Razi Vaccine and Serum Research Institute, Arak for supporting the  
139 research.

### 140 **Author Contributions**

141 Fatemeh Arabkhazaeli: Conceptualization, Data curation, Investigation, Methodology,  
142 Software, Supervision, Writing – original draft, Writing – review & editing. Seyed Davood  
143 Hosseini: Supervision, Validation, Writing – review & editing. Mobina Farrokhnia: Writing –  
144 original draft. Mohammad Khani: Investigation, Writing – original draft.

### 145 **Conflict of Interest**

146 The authors declare that the research was conducted in the absence of any commercial or  
147 financial relationships that could be construed as a potential conflict of interest.

### 148 **Funding**

149 The project was funded under grant number (7405074/8/9), Faculty of Veterinary Medicine,  
150 University of Tehran.

### 151 **Reference**

- 152 1. Dubey JP, Schares G, Ortega-Mora L. Epidemiology and control of neosporosis and  
153 *Neospora caninum*. *Clinical microbiology reviews*. 2007 Apr;20(2):323-67.
- 154 2. Donahose SL, Lindsay SA, Krockenberger M, Phalen D, Šlapeta J. A review of neosporosis  
155 and pathologic findings of *Neospora caninum* infection in wildlife. *International Journal for*  
156 *Parasitology: Parasites and Wildlife*. 2015 Aug 1;4(2):216-38.
- 157 3. Gharekhani J, Yakhchali M, Heidari R. Molecular detection and phylogenetic analysis of  
158 *Neospora caninum* in various hosts from Iran. *Comparative Immunology, Microbiology and*  
159 *Infectious Diseases*. 2022 Jan 1;80:101737.
- 160 4. Evans J, Levesque D, Shelton GD. Canine inflammatory myopathies: a clinicopathologic  
161 review of 200 cases. *Journal of Veterinary Internal Medicine*. 2004 Sep;18(5):679-91.
- 162 5. Dubey JP, Vianna MC, Kwok OC, Hill DE, Miska KB, Tuo W, Velmurugan GV, Conors M,  
163 Jenkins MC. Neosporosis in Beagle dogs: clinical signs, diagnosis, treatment, isolation and genetic  
164 characterization of *Neospora caninum*. *Veterinary parasitology*. 2007 Nov 10;149(3-4):158-66.
- 165 6. Gual I, Campero LM, Hecker YP, Regidor-Cerrillo J, Leunda MR, Odeón AC, Campero CM,  
166 Torioni de Echaide S, Echaide IE, Estein SM, Ortega-Mora LM. Parasitemia and Associated  
167 Immune Response in Pregnant and Non-Pregnant Beef Cows Naturally Infected With *Neospora*  
168 *caninum*. *Frontiers in Veterinary Science*. 2022 Jun 14;9:905271.
- 169 7. Reichel MP, Wahl LC, Ellis JT. Research into *Neospora caninum*—what have we learnt in  
170 the last thirty years?. *Pathogens*. 2020 Jun 23;9(6):505.
- 171 8. Anvari D, Saberi R, Sharif M, Sarvi S, Hosseini SA, Moosazadeh M, Hosseininejad Z,  
172 Chegeni TN, Daryani A. Seroprevalence of *Neospora caninum* infection in dog population  
173 worldwide: a systematic review and meta-analysis. *Acta parasitologica*. 2020 Jun;65:273-90.
- 174 9. Abdoli A, Arbabi M, Pirestani M, Mirzaghavami M, Ghaffarifar F, Dalimi A, Sadraei J.  
175 Molecular assessment of *Neospora caninum* and *Toxoplasma gondii* in hooded crows (*Corvus*  
176 *cornix*) in Tehran, Iran. *Comparative immunology, microbiology and infectious diseases*. 2018 Apr  
177 1;57:69-73.
- 178 10. Nazari N, Shojaee S, Salimi M, Mohebbali M, Ahmadifard N, Hamzavi Y, Zarei Z,  
179 Farahmand-Rad R, Bozorgomid A, Heydarian P. Serological survey of *Neospora caninum* and  
180 *Toxoplasma gondii* co-infection in rodents in Northwestern Iran. *Iranian Journal of Parasitology*.  
181 2020 Apr;15(2):253.

11. Fernández-Escobar M, Millán J, Chirife AD, Ortega-Mora LM, Calero-Bernal R. Molecular survey for cyst-forming coccidia (*Toxoplasma gondii*, *Neospora caninum*, *Sarcocystis* spp.) in Mediterranean periurban micromammals. *Parasitology Research*. 2020 Aug;119(8):2679-86.
12. Medina-Esparza L, Macías L, Ramos-Parra M, Morales-Salinas E, Quezada T, Cruz-Vázquez C. Frequency of infection by *Neospora caninum* in wild rodents associated with dairy farms in Aguascalientes, Mexico. *Veterinary Parasitology*. 2013 Jan 16;191(1-2):11-4.
13. Gharekhani J, Yakhchali M, Berahmat R. *Neospora caninum* infection in Iran (2004–2020): A review. *Journal of Parasitic Diseases*. 2020 Dec;44(4):671-86.
14. Khani M, Arabkhazaeli F, Hosseini SD, Shayan P. Molecular detection of *Neospora caninum* in aborted fetuses of cattle farms in Arak. *Journal of Veterinary Research*. 2018 Dec 73(4):457-463.
15. Meerburg BG, De Craeye S, Dierick K, Kijlstra A. *Neospora caninum* and *Toxoplasma gondii* in brain tissue of feral rodents and insectivores caught on farms in the Netherlands. *Veterinary parasitology*. 2012 Mar 23;184(2-4):317-20.
16. Jenkins MC, Parker C, Hill D, Pinckney RD, Dyer R, Dubey JP. *Neospora caninum* detected in feral rodents. *Veterinary Parasitology*. 2007 Jan 31;143(2):161-5.
17. Okeoma CM, Williamson NB, Pomroy WE, Stowell KM, Gillespie L. The use of PCR to detect *Neospora caninum* DNA in the blood of naturally infected cows. *Veterinary parasitology*. 2004 Aug 6;122(4):307-15.
18. Khan A, Shaik JS, Sikorski P, Dubey JP, Grigg ME. Neosporosis: an overview of its molecular epidemiology and pathogenesis. *Engineering*. 2020 Feb 1;6(1):10-9.
19. Ferroglio E, Pasino MA, Romano A, Grande D, Pregel P, Trisciuglio A. Evidence of *Neospora caninum* DNA in wild rodents. *Veterinary Parasitology*. 2007 Sep 30;148(3-4):346-9.
20. Dellarupe A, Fite B, Pardini L, Campero LM, Bernstein M, Robles MD, Moré G, Venturini MC, Unzaga JM. *Toxoplasma gondii* and *Neospora caninum* infections in synanthropic rodents from Argentina. *Revista Brasileira de Parasitologia Veterinária*. 2019 Jan;28:113-8.
21. Hůrková-Hofmannová L, Qablan MA, Jurankova J, Modrý D, Pialek J. A survey of *Toxoplasma gondii* and *Neospora caninum* infecting house mice from a hybrid zone. *Journal of parasitology*. 2014 Feb 1;100(1):139-41.
22. Muradian V, Ferreira LR, Lopes EG, de Oliveira Esmerini P, de Jesus Pena HF, Soares RM, Gennari SM. A survey of *Neospora caninum* and *Toxoplasma gondii* infection in urban rodents from Brazil. *Journal of Parasitology*. 2012 Feb 1;98(1):128-34.
23. Nazari N, Shojaee S, Mohebbali M, Teimouri A, Ghadiri K, Raeghi S, Shiee MR, Azarakhsh Y, Bozorgomid A. *Toxoplasma gondii* and *Neospora caninum* in brain tissue of rodents in North-West Iran. *Veterinary Medicine: Research and Reports*. 2019 Dec 20:223-7.
24. Macháčová T, Ajzenberg D, Žáková A, Sedlák K, Bártová E. *Toxoplasma gondii* and *Neospora caninum* in wild small mammals: seroprevalence, DNA detection and genotyping. *Veterinary Parasitology*. 2016 Jun 15;223:88-90.
25. Japa O, Morand S, Karnchanabanthoeng A, Chaisiri K, Ribas A. Detection of *Neospora caninum* (*Toxoplasmatidae*) in wild small mammals from Thailand. *Folia Parasitologica*. 2018 Jan 1;65.
26. Hamzavi Y, Salimi Y, Ahmadi M, Adimi P, Falahi S, Bozorgomid A. Global prevalence of *Neospora caninum* in rodents: A systematic review and meta-analysis. *Veterinary Medicine and Science*. 2023 Sep;9(5):2192-200.
27. Coombs RS. Immediate IFN $\gamma$  production determines host compatibility differences between *Toxoplasma gondii* and *Neospora caninum* in mice (Doctoral dissertation, University of Pittsburgh).